



Article

Diversity of ribosomes at the level of rRNA variation associated with human health and disease

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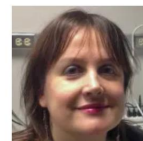
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Maria Barna - Associate Professor of Genetics



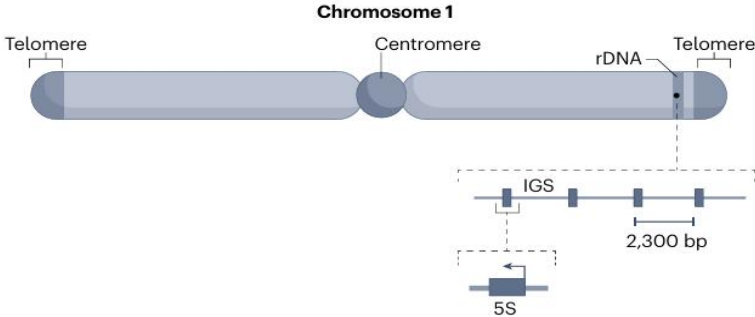
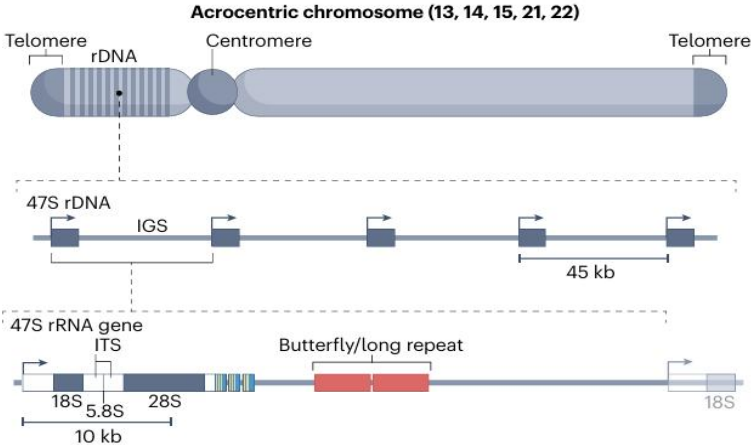
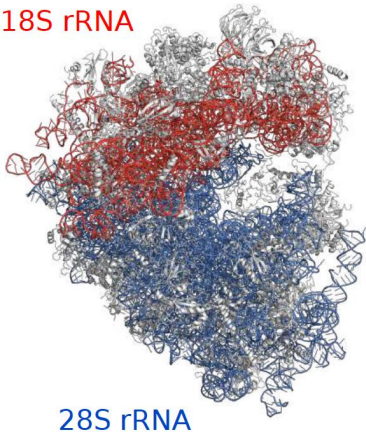
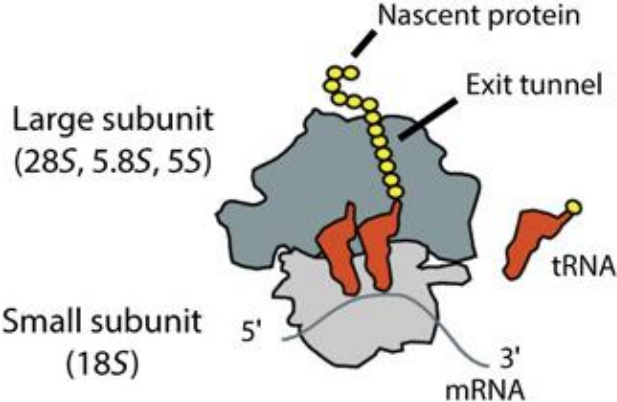
Bio-X Affiliated Faculty

[Stanford Profile](#) | [Dr. Barna's Lab Homepage](#)

Dr. Maria Barna's lab studies how intricate control of gene expression and cell signaling is regulated on a minute-by-minute basis to give rise to the remarkable diversity of cell types and tissue morphology that form the living blueprints of developing organisms. This research aims to add a new dimension to our understanding of how cells “know” where to go, when to move and differentiate by employing novel technologies that probe these questions at a highly molecular and nanoscale level. Work in the Barna lab is presently split into two main research efforts. The first is investigating “specialized ribosomes” and mRNA translation in control of gene expression genome-wide in space and time during development. This research is opening a new field of study in which fundamental aspects of gene regulation are controlled by ribosomes harboring a unique activity that “select” for specific mRNAs to translate by virtue of unique RNA regulons embedded within 5'UTRs. The second research effort is centered on employing state-of-the-art live cell imaging to visualize cell signaling and cellular control of organogenesis. This research has led to the realization of a novel means of cell-cell communication dependent on a dense network of actin-based cellular extension within developing organs that interconnect and facilitate the precise transmission of molecular information between cells.

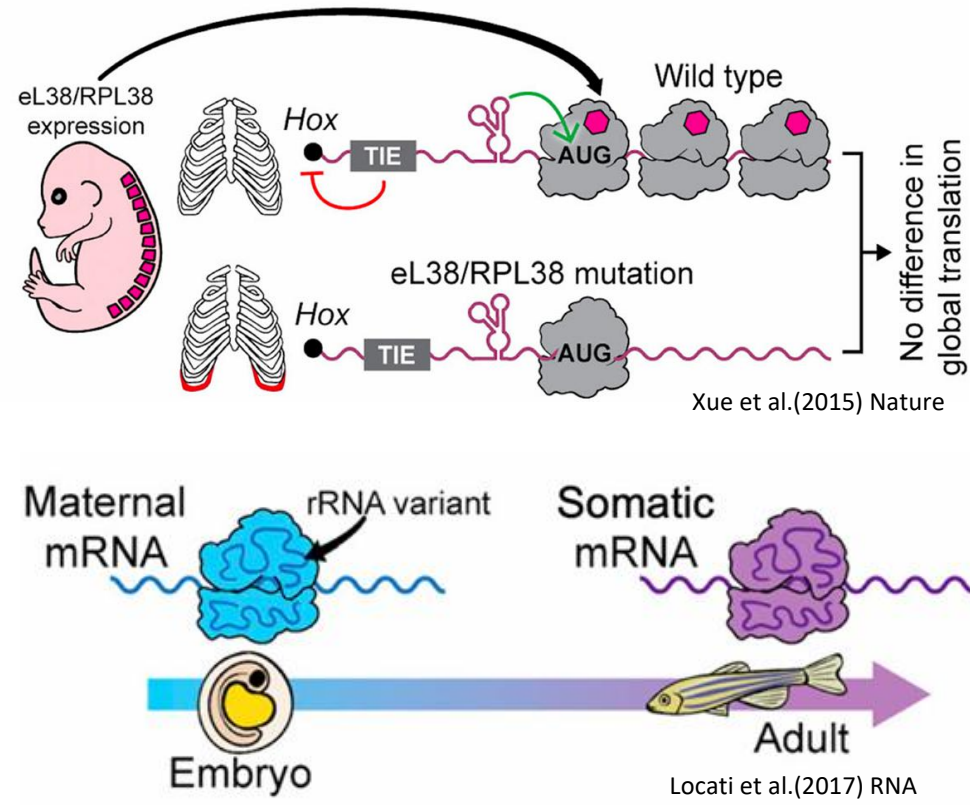
Present via Lei, NIE

Background



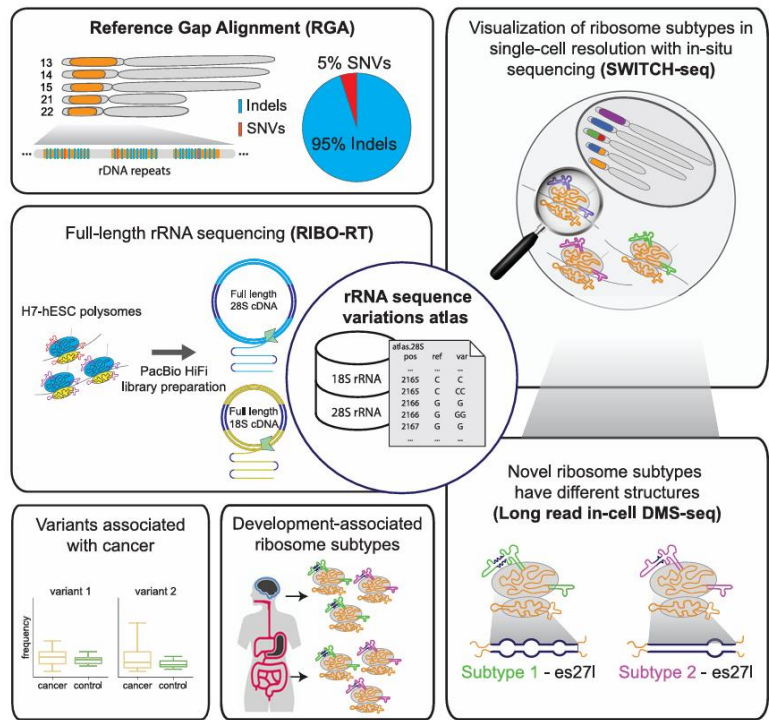
Background

ribosomes are heterogeneous

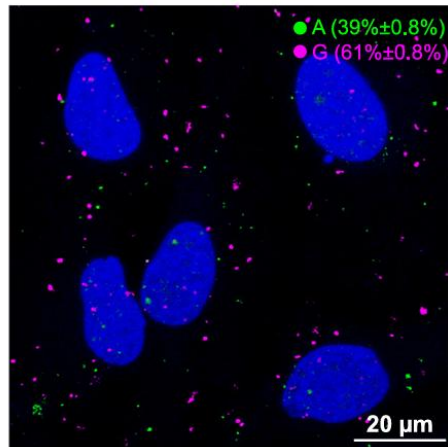
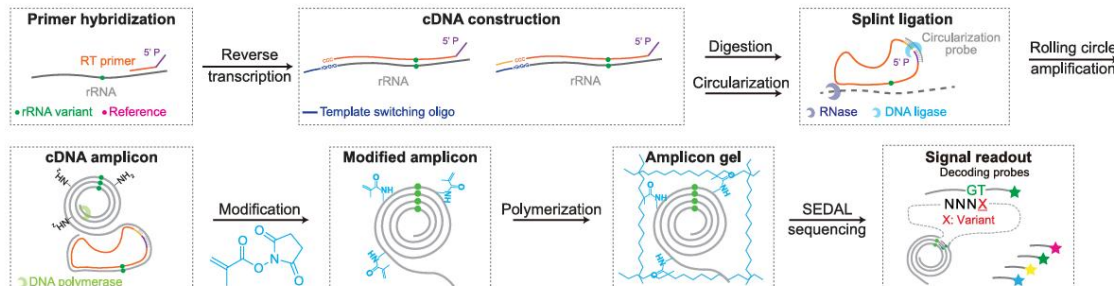
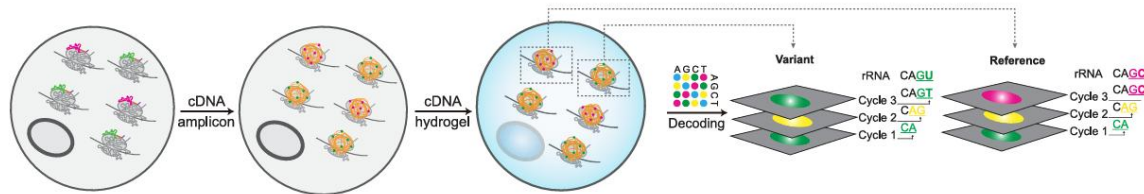


Inconsistent variant calling

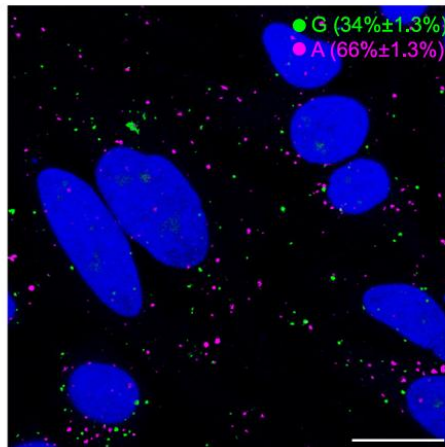
Study	Dataset	Technology	SNVs	Indels
Parks et al. (Sci Adv, 2018)	1KGP	Illumina short-read	97.3%	2.7%
Fan et al. (RNA, 2022)	1KGP	Illumina short-read	80.8%	19.2%



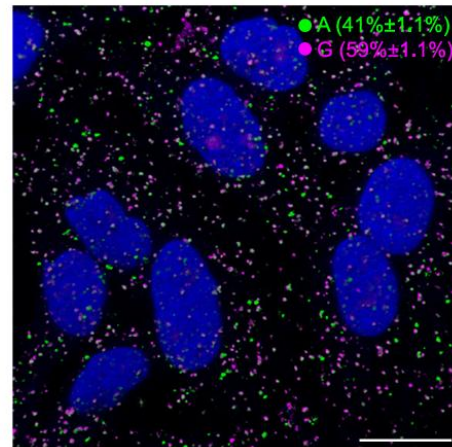
The co-expressed pattern visualized by *in situ* sequencing (SWITCH-seq)



28S:h11 - position 60



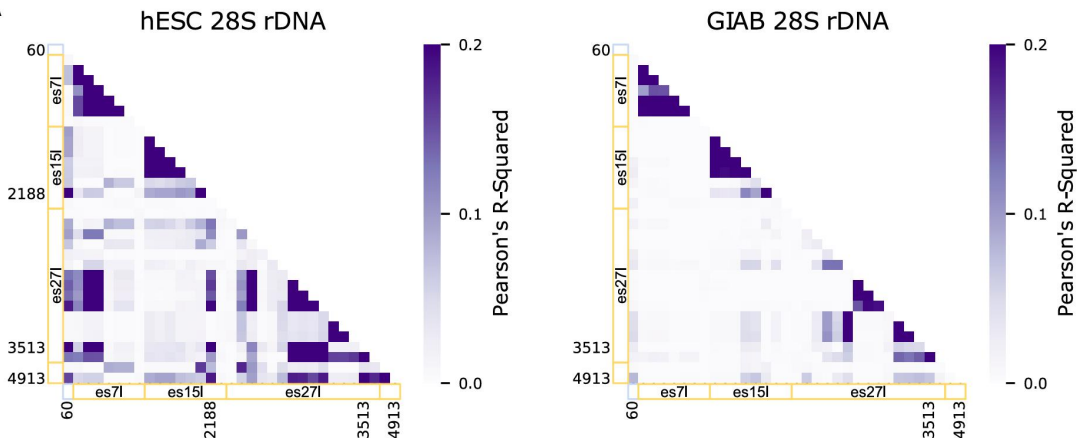
es27I - position 3040



es27I - position 3491

Three variants chosen for haplotyping the 28S rDNA

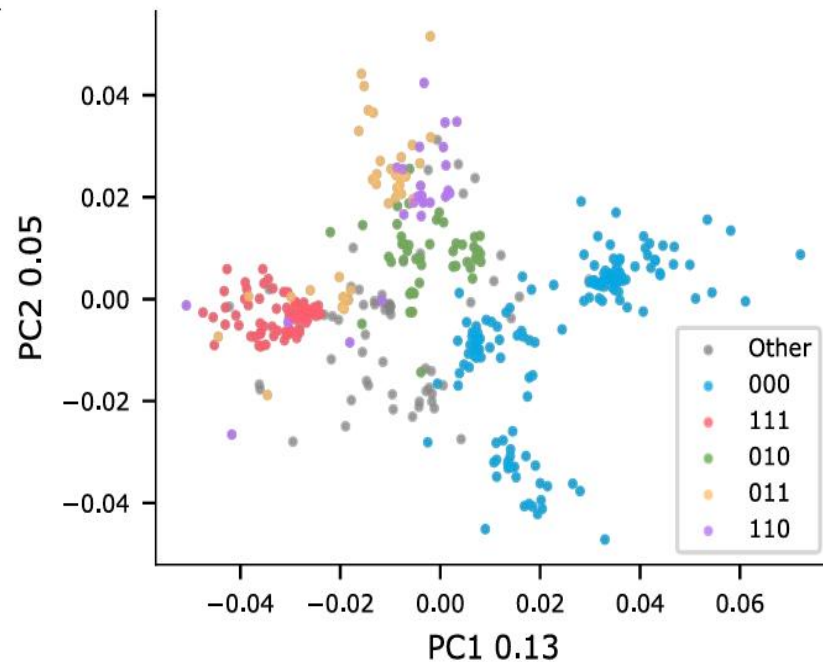
Association of Variants



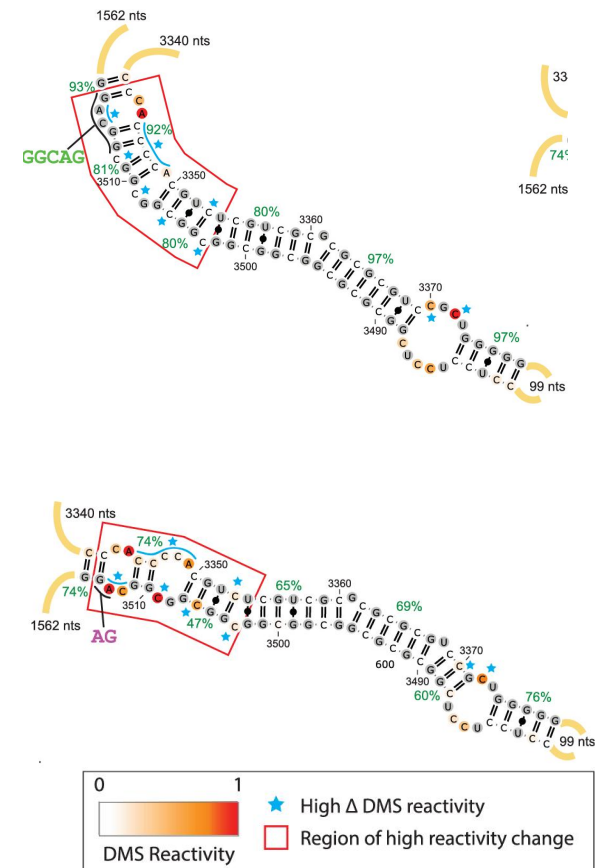
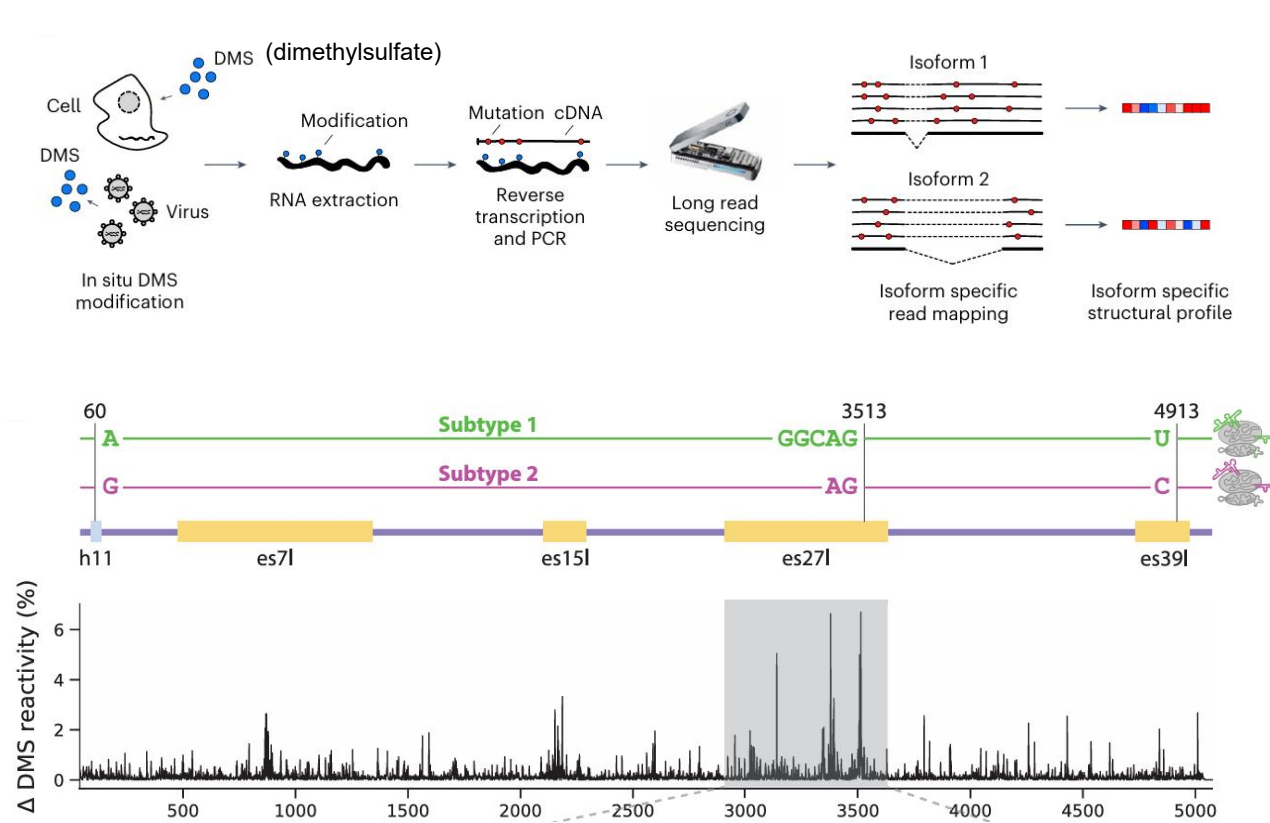
28S position	Digit	Nucleotide
60	0	G
	1	A
3513	0	AG
	1	GGCAG
	2	GG
4913	0	C
	1	T

28S pairwise similarity in hESC

hESC complete 28S

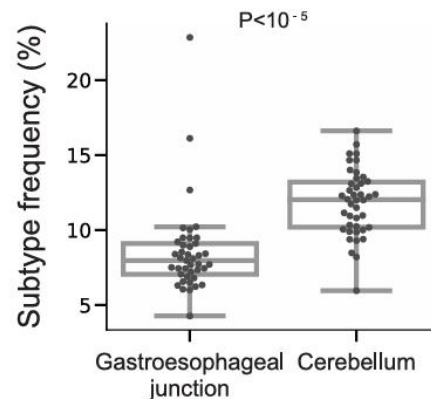
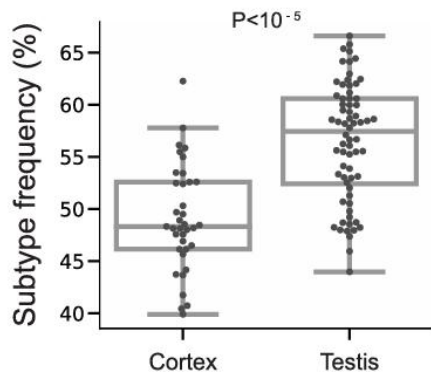
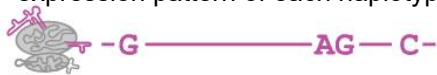


Variants leading to secondary structure difference

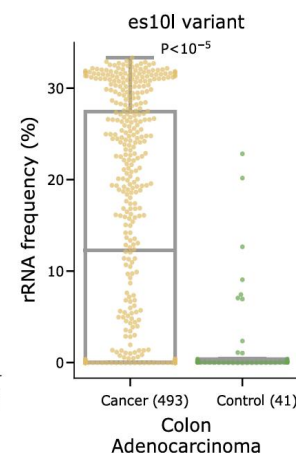
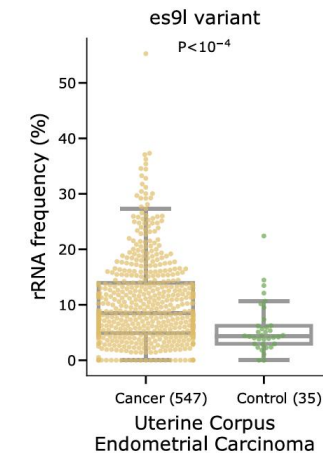
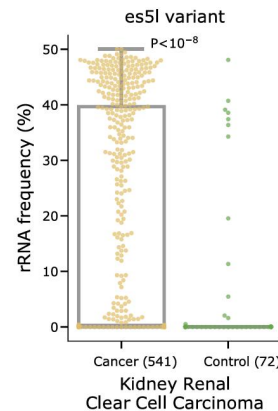
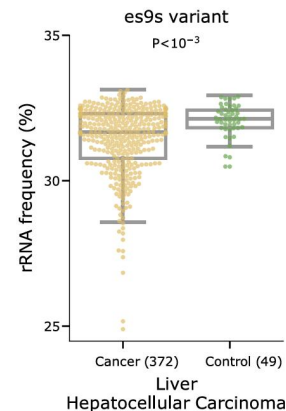


The relative abundance of rRNA variants in expression data

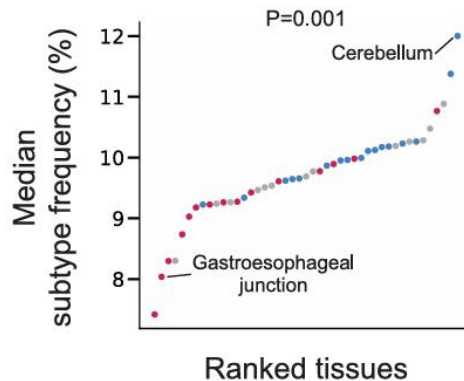
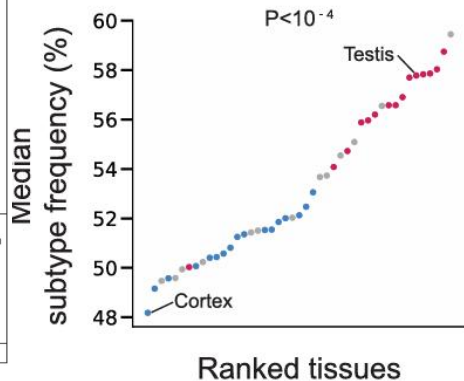
expression pattern of each haplotype is tissue specific



Some rRNA variants are cancer specific



• Ectoderm derived tissues	
Amygdala	
Anterior cingulate cortex	
Caudate nucleus	
Cerebellar hemisphere (Snap frozen)	
Cerebellum (Fixed tissue)	
Cortex (Fixed tissue)	
Frontal cortex (Snap frozen)	
Hippocampus	
Hypothalamus	
Nucleus accumbens	
Pulvinar	
Spinal cord	
Substantia nigra	
Skin - sun exposed	
Skin - not sun exposed	
• Endoderm derived tissues	
Esophagus mucosa	Transverse colon
Esophagus muscularis	Thyroid
Gastroesophageal junction	Testis
Liver	Vagina
Pancreas	Ovary
Sigmoid colon	Uterus
Stomach	Prostate
Terminal ileum	
• Other tissues	



Conclusions

- New methods (RGA, RIBO-RT, SWITCH-seq) enable comprehensive study of rRNA variations.
- The rRNA subtypes show structural differences and tissue-specific expression.
- Some rRNA variants are linked to cancer.

Future Directions

- Can the subtype be determined just by one variant not a combination of variants
- Does the subtype frequency differences exist in different population, or in people of different ages.